

## CURRICULUM VITAE

**Name:** Andreia Albuquerque Wendt

**Position title:** Advanced Postdoctoral Researcher / Marie Curie Global Fellow

**Affiliations:** Parasite Chemotherapy Unit, Swiss TPH (CH); Faculty of Sciences, Charles University Prague (CZ); GHTM, IHMT-NOVA University (PT)

**ORCID:** <https://orcid.org/0000-0001-5794-5417>

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## PERSONAL STATEMENT

Dr. Wendt graduated in 2018 from the Medizinische Hochschule Hannover, with a PhD in Molecular Medicine. During her first postdoc in the laboratory of Prof. Eva Gluenz, she focused on the role of the *Leishmania mexicana* transportome across its life cycle using BAR-seq methodologies. Recently, she started a second postdoctoral project focusing on the development of anti-*Leishmania* transmission blocking sand fly sugar baits supported by Prof. Pascal Maeser and Prof. Petr Volf. Dr. Wendt's research aims to explore the intricate details of host-pathogen-vector interactions in *Leishmania* infections and ultimately harvest basic information for the development of control strategies contributing for the eradication of leishmaniasis.

## RESEARCH INTERESTS

Molecular & Cellular Parasitology, Vector-Parasite-Host interactions, Drug development, VBD control.

## EDUCATION

**26/01/2018 Doctor of Philosophy (PhD), Molecular Medicine**, Medizinische Hochschule Hannover, DE

Thesis title: Glycosylation in Apicomplexan Parasites. Supervisor: Prof. Dr. Françoise Routier

**11/12/2013 Master (MSc) of Biomedical Sciences - Molecular Biology and International Tropical Medicine**, Instituto de Higiene e Medicina Tropical – Universidade Nova de Lisboa, PT

Thesis title: New species of *Leishmania* and hybrids – implications in the pathogenesis of infection.

Supervisors: Prof. Dr. Lenea Campino and Dr. Sofia Cortes

**23/09/2011 Bachelor (BSc) in Biochemistry**, Faculdade de Ciências e Tecnologias, Universidade NOVA de Lisboa, Monte da Caparica, PT

Thesis title: Spectroscopic characterization of chalcone (1E, 4E)-1,5-BIS(4-hidroxyfenyl)penta-1,4-diene-3-one. Supervisor: Prof. Dr. César Laia

**15/09/2010 Bachelor (BSc) in Applied Chemistry - Biotechnology**, Faculdade de Ciências e Tecnologias, Universidade NOVA de Lisboa, Monte da Caparica, PT

Thesis title: Study of in vitro biological behavior of parental and hybrid strains of *Leishmania*. Supervisors: Prof. Dr. Lenea Campino and Dr. Sofia Cortes

## PROFESSIONAL EXPERIENCE

**Since 01/04/2025 Guest Assistant Professor**, IHMT-UNL, Lisbon, Portugal

Description: Responsibility for the teaching at IHMT, collaboration in research conducted at GHTM.

**Since 01/11/2024 Advanced postdoctoral researcher (MSCA GF Fellow)**, Swiss TPH, Basel, Switzerland & Faculty of Sciences, Charles University, Prague, Czech Republic

Description: Research responsibility for the development of a sand fly colony at the insectary of Swiss TPH, to capacitate infection and chemotherapy experiments using *Leishmania* sp.. Generate *Leishmania* sp. genetically modified cell lines containing defined fluorescent tags and unique nucleotide barcodes, enabling NGS based medium to high-throughput compound screening of parasiticide agents in sand flies.

**01/11/2023 – 31/10/2024 Advanced postdoctoral researcher**, Institute of Cell Biology, University of Bern, Bern, Switzerland

Description: Research responsibility for producing DNA constructs for the generation of selected knockout, add-back and tagged *L. mexicana* cell and phenotypic analysis in vitro. Drug screenings in *L. mexicana* and *L. donovani* axenic promastigotes and amastigotes, as well as intracellular amastigotes, using iMAC models,

in collaboration with PCU (Swiss TPH). Data analysis, manuscript preparation and outreach. Shared responsibility for supervision of undergraduate and graduate students.

**01/09/2020 – 30/08/2023 Postdoctoral researcher**, Wellcome Centre for Integrative Parasitology, Institute of Infection & Immunity, College of Medical Veterinary & Life Sciences, University of Glasgow, Glasgow, UK

Description: Research responsibility for producing DNA constructs for the generation of a knockout library (II) of more than 300 individually barcoded mutants and phenotypic analysis of this library in vivo (macrophages, mice and sand flies). Data analysis, manuscript preparation and outreach. Shared responsibility for supervision of undergraduate and graduate students.

**01/09/2018 – 30/08/2020 Postdoctoral researcher (MSCA IF Fellow)**, Sir William Dunn School of Pathology, University of Oxford, United Kingdom

Description: Research responsibility for producing DNA constructs for the generation of a knockout library (I) of more than 300 individually barcoded mutants and phenotypic analysis of this library in vitro (promastigotes). Data analysis, manuscript preparation and outreach. Shared responsibility for supervision of undergraduate and graduate students.

**01/02/2018 – 30/08/2018 Postdoctoral researcher**, Instituts für Klinische Biochemie, Medizinische Hochschule Hannover, DE

Description: Following graduate studies, research responsibility for maintaining various cell lines including Schneider insect cells, CHO, Vero and HFF cells as well as *Toxoplasma gondii* parental and mutant cell lines. Additional tasks included the expression of various recombinant proteins in eukaryotic heterologous systems, purification, and mass spectrometry analysis. Data analysis, manuscript preparation and outreach. Shared responsibility for supervision of undergraduate and graduate students.

**01/06/2011 – 30/06/2014 Research Assistant**, Unidade de Parasitologia Médica, Instituto de Higiene e Medicina Tropical – Universidade NOVA de Lisboa, Lisbon, PT

Description: Research responsibility for processing biological samples from dogs and murine model for the isolation, detection and typing of new *Leishmania* strains, developing highly sensitive diagnostic tools for detection of asymptomatic parasitemia, as well as development and testing of leishmanicidal compounds. Shared responsibility for progress reporting to National Science Foundation. Data analysis, manuscript preparation and outreach. Shared responsibility for team management including supervision of undergraduate and graduate students. During this period, she participated in several national and international projects, namely the “New *Leishmania* species and hybrids – risk of introduction and implications to the pathogeny of infection” (PTDC/CVT/112371/2009), financed by Portuguese foundation for Science and Technology (FCT), and “Biology and control of vector-borne infections in Europe- EDEN-next”, financed by EU/FP7 and also “Network for identification of biomarkers potentially useful in discriminating the different clinical outcomes of *Leishmania infantum* infection”, CYTED-Brazil.

## RESEARCH MONOGRAPHS

1. **A. Albuquerque**. 2018. Glycosylation in Apicomplexan Parasites. Doctor of Philosophy (PhD) thesis in Molecular Medicine. Medizinische Hochschule Hannover
2. **A. Albuquerque**. 2013. New species of *Leishmania* and hybrids – implications in the pathogenesis of infection. Master (MSc) thesis in Biomedical Sciences: Molecular biology and International Tropical Medicine. Instituto de Higiene e Medicina Tropical – Universidade NOVA de Lisboa
3. **A. Albuquerque**. 2010. Study of in vitro biological behavior of parental and hybrid strains of *Leishmania*. Bachelor (BSc) thesis in Applied Chemistry – Biotechnology. Faculdade de Ciências e Tecnologias, Universidade NOVA de Lisboa

## PUBLISHED JOURNAL ARTICLES (\*co-first)

1. **A. Albuquerque-Wendt**, C. McCoy, R. Neish, U. Dobramysl, Ç. Alagöz, T. Beneke, S.A. Cowley, K. Crouch, R.J. Wheeler, J.C. Mottram and E. Gluenz. TransLeish: Identification of membrane transporters

- essential for survival of intracellular *Leishmania* parasites in a systematic gene deletion screen. *Nature Communications*, **2025**. 16, 299. Doi: 10.1038/s41467-024-55538-7.  
*Role*: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Supervision, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
2. T. Beneke, R. Neish, C. M. C. Catta-Preta, J. Smith, J. Valli, C. McCoy **A. Albuquerque-Wendt**, J.C. Mottram and E. Gluenz. IFT and BBSome proteins are required for *Leishmania mexicana* pathogenicity, but flagellar motility is dispensable. *bioRxiv*, **2024**. Doi: 10.1101/2024.09.13.612850.  
*Role*: Investigation, Writing – Review & Editing.
  3. C. Ricce Espada, J. C. Quilles Jr, **A. Albuquerque-Wendt**, M. C. Cruz, T. Beneke, L. Lorenzon, E. Gluenz, A. K. Cruz, S. R. B. Uliana. Effective genome editing in *Leishmania (Viannia) braziliensis* stably expressing Cas9 and T7 RNA Polymerase. *Frontiers in Cellular and Infection Microbiology*, **2021**. Doi: 10.3389/fcimb.2021.772311.  
*Role*: Conceptualization, Supervision, Data Curation, Formal Analysis, Writing – Review & Editing.
  4. C. Ricce Espada, **A. Albuquerque-Wendt**, V. Hornillos, E. Gluenz, A.C. Coelho, S. Uliana. Ros3 (Lem3p/CDC50) gene dosage is implicated in miltefosine susceptibility in *Leishmania (Viannia) braziliensis* clinical isolates and in *Leishmania (Leishmania) major*. *ACS Infectious Diseases*, **2021**. 7(4): 849-858. Doi: 10.1021/acsinfecdis.0c00857.  
*Role*: Conceptualization, Supervision, Data Curation, Formal Analysis, Writing – Review & Editing.
  5. **A. Albuquerque-Wendt**, D. Jacot, N. Pacheco, C. Seegers, P. Zarnovican, D. Soldati-Favre, H. Bakker, F. H. Routier. C-Mannosylation of *Toxoplasma gondii* proteins promotes attachment to host cells and parasite virulence. *Journal of Biological Chemistry*, **2020**, 295(4): 1066-1076. Doi: 10.1074/jbc.RA119.010590.  
*Role*: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
  6. **A. Albuquerque-Wendt**, H. J. Hütte, F. F. R. F. Buettner, H. Routier, H. Bakker. Membrane Topological Model of Glycosyltransferases of the GT-C Superfamily. *International journal of molecular sciences*, **2019**, 20(19): E4842. Doi: 10.3390/ijms20194842.  
*Role*: Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
  7. G. Bandini\*, **A. Albuquerque-Wendt\***, J. Hegermann, J. Samuelson, F. H. Routier. Protein O- and C-glycosylation pathways in *Plasmodium falciparum* and *Toxoplasma gondii*. *Parasitology*, **2019**, 146(14): 1755-1766. Doi: 10.1017/S0031182019000040.  
*Role*: Conceptualization, Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
  8. S. Cortes\*, **A. Albuquerque-Wendt\***, C. Maia, M. Carvalho, L. Lima, L. A. R. Freitas, W. L. C. dos-Santos, L. Campino. Elucidating *in vitro* and *in vivo* behaviour of *L. infantum*/*L. major* natural hybrids. *Parasitology*, **2019**, 146(5): 580-587. Doi: 10.1017/S0031182018001993.  
*Role*: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
  9. C. Hoppe, **A. Albuquerque-Wendt**, A. Shcherbakova, F. F. R. Buettner, L. Izquierdo, H. Bakker, F. H. Routier. Apicomplexan C-mannosyltransferases modify thrombospondin type I containing adhesins of the TRAP family. *Glycobiology*, **2018**, 28(5): 333-343. Doi: 10.1093/glycob/cwy013.  
*Role*: Investigation, Writing – Review & Editing.
  10. **A. Albuquerque**, L. Campino, L. Cardoso, S. Cortes. Evaluation of four molecular methods to detect *Leishmania* infection in dogs. *Parasites & Vectors*, **2017**, 10(1): 57. Doi: 10.1186/s13071-017-2002-2.  
*Role*: Conceptualization, Methodology, Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.
  11. S. Cortes\*, **A. Albuquerque\***, L. Cabral, L. Lopes, L. Campino, M. L. Cristiano. *In vitro* susceptibility of *Leishmania infantum* to Artemisinin Derivatives and selected Trioxolanes. *Antimicrobial Agents and Chemotherapy*, **2015**, 59(8): 5032-5. Doi: 10.1128/AAC.00298-15.

*Role:* Conceptualization, Methodology, Supervision, Project administration, Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.

12. **A. Albuquerque**, L. Campino, S. Cortes. 2012. In vitro biological behavior of *Leishmania infantum/L. major* hybrid strains from Mediterranean basin, Portugal. In XI European Multicolloquium of Parasitology, V. Cosma (Ed.), Medimond International Proceedings, Bolonha, pp. 5-9.

*Role:* Validation, Formal analysis, Investigation, Data Curation, Writing – Original Draft, Writing – Review & Editing, Visualization.

*Role definitions as per defined by [CRediT](#)*

## ORAL COMMUNICATIONS (Last 10 years)

1. **A. Albuquerque-Wendt**. 2024. A transportome-wide phenotypic screen identifies membrane transporter proteins essential for parasite fitness in the vertebrate and invertebrate hosts. Biozentrum Universität Würzburg, Germany (*by invitation*)
2. **A. Albuquerque-Wendt**. 2024. Utilising bar-seq strategies for the identification of *Leishmania* genes essential for survival in the vertebrate and invertebrate hosts. “Future of Parasitology”, Institute Pasteur, Paris, France (*by invitation*)
3. **A. Albuquerque-Wendt**. 2024. TransLeish: a systematic gene deletion screen identifies an essential role for the *Leishmania* V-ATPase in life cycle progression. XXXV Molecular Parasitology Meeting, Woods Hole, MA, United States of America
4. **A. Albuquerque-Wendt**. 2024. Characterisation of *L. mexicana* ABCI3 knockout mutants in vitro and in vivo. 40<sup>th</sup> Swiss Trypanosomatid Meeting, Les Diablerets, Switzerland
5. **A. Albuquerque-Wendt**. 2023. Science in Nepal: Collaboration Opportunities”. GHTM Sessions. Instituto de Higiene e Medicina Tropical – Universidade NOVA de Lisboa, Lisbon, Portugal (*by invitation*)
6. **A. Albuquerque-Wendt**. 2023. Transporters be Gone: CRISPR/Cas9 knockout screen of the transportome of *Leishmania mexicana*. Charles Prague University, Prague, Czech Republic (*by invitation*)
7. **A. Albuquerque-Wendt**, T. Beneke, C. McCoy, S. Cowley, R. Neish, J. Mottram, E. Gluenz. 2023. Fitness screen of *Leishmania* membrane transporters. 39<sup>th</sup> Swiss Trypanosomatid Meeting, Leysin, Switzerland
8. **A. Albuquerque-Wendt**, T. Beneke, C. McCoy, S. Cowley, R. Neish, J. Mottram, E. Gluenz. 2022. CRISPR/Cas9 genetic screen in *Leishmania mexicana* identifies membrane transporters important for parasite survival in the mammalian host. 3rd International Caparica Conference on Leishmaniasis 2022, Caparica, Portugal
9. **A. Albuquerque-Wendt**, T. Beneke, C. McCoy, S. Cowley, R. Neish, J. Mottram, E. Gluenz. 2022. Identification of transporters proteins important for in vitro and in vivo parasite survival in *Leishmania mexicana*. Swiss TPH, Basel, Switzerland (*by invitation*)
10. **A. Albuquerque-Wendt**, T. Beneke, C. McCoy, S. Cowley, R. Neish, J. Mottram, E. Gluenz. 2022. CRISPR/Cas9 transportome genetic screen in *Leishmania mexicana* identifies membrane transporters important for parasite survival. WorldLeish7, Cartagena, Colombia
11. **A. Albuquerque-Wendt**. 2018. A Trypanosomatidae-Apicomplexan playground: A sweet and bumpy rollercoaster. Faculdade de Ciências e Tecnologias, Universidade NOVA de Lisboa, Lisbon, Portugal (*by invitation*)
12. **A. Albuquerque-Wendt**. 2018. A Trypanosomatidae-Apicomplexan playground: A sweet and bumpy rollercoaster. Instituto de Higiene e Medicina Tropical – Universidade NOVA de Lisboa, Lisbon, Portugal (*by invitation*)
13. **A. Albuquerque**, C. Hoppe, A. Shcherbakova, L. Izquierdo, F. Buettner, H. Bakker, F.H. Routier. 2017. C-mannosylation of *Toxoplasma gondii* Thrombospondin Type 1 Repeats. Glycobiology Gordon Research Seminar, Ventura, CA, USA
14. **A. Albuquerque**. 2016. Mind sucking Parasites – Part II – The Empire strikes back. Science Slam, Leibniz Universität, Hannover, Germany
15. **A. Albuquerque**. 2016. Glycosylation of *Toxoplasma gondii* Thrombospondin Type 1 Repeats. GlycoPar symposium. Liverpool School of Tropical Medicine, United Kingdom

16. **A. Albuquerque**. 2015. Mind sucking Parasites. Science Slam. Leibniz Universität, Hannover, Germany
17. S. Cortes, **A. Albuquerque**, L. Cabral, L. Lopes, M.L. Cristiano, L. Campino. 2015. Potential of Artemisinin derivatives and trioxolanes as anti-*Leishmania* chemotherapy. National Symposium on Zoonoses Research, Berlin, Germany

## POSTER PRESENTATIONS (Last 10 years)

1. **A. Albuquerque-Wendt**, B. Vojtková, T. Bečvář, J. Sadlova, P. Volf. 2025. LEISHBLOCK: Exploring an antileishmanial drug toolbox for blocking pathogen transmission in the sand fly vector. Joint Parasitology SSTMP, DGP, BSP Spring Meeting, Würzburg, Germany
2. **A. Albuquerque-Wendt**, B. Vojtková, T. Bečvář, C. McCoy, R. Neish, U. Dobramysl, C. Alagöz, T. Beneke, S.A. Cowley, K. Crouch, J. Sadlova, P. Volf, R.J. Wheeler, J.C. Mottram, E. Gluenz. 2025. Utilising bar-seq strategies for the identification of *Leishmania* genes essential for survival in the sand fly vector. SVEG annual scientific meeting, Lausanne, Switzerland
3. **A. Albuquerque-Wendt**, T. Beneke, C. McCoy, S. Cowley, R. Neish, J. Mottram, E. Gluenz. 2022. High-throughput Genetic Screen of the *Leishmania mexicana* transportome highlights transporter proteins important for parasite fitness in vitro and in vivo. Membrane proteins workshop & SMALP, Birmingham, United Kingdom
4. **A. Albuquerque-Wendt**, T. Beneke, C. C. Espada, McCoy, E. Gluenz. 2020. TransLeishlon: Dissecting the role of membrane transporter proteins in *Leishmania mexicana*. 2nd International Caparica Conference on Leishmaniasis 2020, Caparica, Portugal (on-line)
5. **A. Albuquerque-Wendt**, E. Gluenz. 2019. TransLeishlon: *Leishmania* Transportome dissection using CRISPR-Cas9. Sir William Dunn School of Pathology Postdoc Symposium, Oxford, United Kingdom
6. **A. Albuquerque-Wendt**, C. Hoppe, A. Shcherbakova, L. Izquierdo, F. Buettner, H. Bakker, F. Routier. 2018. Apicomplexan C-mannosyltransferases modify adhesins of the TRAP family. BSP Spring Meeting. Aberystwyth, United Kingdom
7. **A. Albuquerque**, C. Hoppe, A. Shcherbakova, L. Izquierdo, F. Buettner, H. Bakker, F. Routier. 2017. C-mannosylation of Apicomplexan TSR containing adhesins. TOXO14. Tomar, Portugal
8. **A. Albuquerque**, L. Cardoso, L. Campino, S. Cortes. 2017. Comparative evaluation of molecular methods to detect *Leishmania* DNA in dogs. BSP Spring Meeting. Dundee, United Kingdom
9. **A. Albuquerque**, C. Hoppe, A. Shcherbakova, L. Izquierdo, F. Buettner, H. Bakker, F. Routier. 2017. C-mannosylation of *Toxoplasma gondii* Thrombospondin Type 1 Repeats. Glycobiology Gordon Research Conference. Ventura, California, United States of America
10. **A. Albuquerque**. Targeting C-mannosylation in Apicomplexan parasites. 2015. BREATH Summer School. International House Sonnenberg, Harz, Germany
11. **A. Albuquerque**, S. Cortes, L. Cabral, L. Lopes, M.L. Cristiano, L. Campino. 2015. Antileishmanial activity of Artemisinin derivatives and selected Trioxolanes. BSP Autumn Meeting – One Health: parasites and beyond. London, United Kingdom

## RESEARCH FUNDING

1. **Marie Skłodowska-Curie Global Fellowship**. Start date: 01.11.2024. End date: 31.10.2027. Funder: Marie Skłodowska-Curie Actions (Horizon Europe). Title: LEISHBLOCK: Exploring an antileishmanial drug toolbox for blocking pathogen transmission in the sand fly vector. Total amount: €309,768.48. Role: Postdoctoral fellow (Awardee).
2. **Wellcome Institutional Strategic Support Fund (ISSF) Feasibility Award**. Start date: 01.11.2022 (4 months). Title: Applying genomics and proteomics for characterisation of metabolic pathways of an *L. mexicana* ABCI3 knock-out cell line. Total: £5,061 (~ €5,930). Role: Leading postdoctoral researcher.
3. **PARSUK-Bilateral Research Fund**. Start date: 01.09.2022 (4 months). Funder: Portuguese Association of Researchers and Students in the UK (PARSUK). Title: LINFTAG – Adapting CRISPR/Cas9 protein tagging in *Leishmania infantum* Portuguese strains. Total amount: €3,000. Role: Leading postdoctoral researcher.

4. **PARSUK-Xperience Scholarship.** Start date: 13.07.2021 (5 weeks). Funder: Portuguese Association of Researchers and Students in the UK (PARSUK). Title: Defining the localisation of calcium transporter proteins in *Leishmania mexicana* parasites. Total amount: €250 (supervisor incentive), €1,500 (supervisee scholarship). Role: Leading postdoctoral researcher.
5. **SULSA ECR Development Award.** Start date: 12.02.2021 (15 months). Funder: Scottish Universities Life Sciences Alliance (SULSA). Title: Essential calcium transporters in *Leishmania*: Biochemical characterisation of *L. mexicana* SERCA and LETM1. Total amount: £4,000 (~ €4,687). Role: Leading postdoctoral researcher.
6. **Marie Skłodowska-Curie Individual Fellowship.** Start date: 01.09.2018. End date: 31.08.2020. Funder: Marie Skłodowska-Curie Actions (European Union FP7). Title: transLEISHion: A targeted knockout screen for identification of Leishmania membrane transporters required for infection of macrophages. Total amount: €195,454. Role: Postdoctoral fellow (Awardee).

#### OTHER AWARDS (excluding travel awards)

1. Funder: Biochemical Society. Total amount: £500 (~ €585). (2023) “Scientific Outreach Grant Further Funding”, supporting the development of the activities during the seminar and workshop at RIBB, Nepal.
2. Funder: Biochemical Society. Total amount: £1000 (~ €1,171). (2023) “Scientific Outreach Grant”, supporting the development of the public engagement activities during the Glasgow Science Festival, United Kingdom.
3. Funder: British Society for Parasitology (BSP). International Training and Fieldwork Award. Total amount: £1,500 (~ €1,757). (2023) Supporting the development of the activities during the seminar and workshop at RIBB, Nepal.
4. Funder: Generon. Best poster presentation. Total amount: £50 (~ €58) (2022). European SMALP, Birmingham, UK.
5. Funder: Anglo-Portuguese Society. Total amount: £500 (~ €585) (2022) Best research pitch prize awarded to the best (out of 10) 3-minute research pitch at the PARSUK 15th LUSO 2022 conference, Ismaili Centre, London, UK.
6. Funder: Wellcome Centre for Integrative Parasitology. Best oral presentation. (2022) Wellcome Centre for Integrative Parasitology/ISAB retreat, Glasgow, UK.
7. Funder: PROTEOMASS Scientific Society. Excellent Shotgun Communication Prize. (2020). 2nd International Caparica Conference on Leishmaniasis, Caparica – Portugal.
8. Funder: Burroughs Wellcome Fund. C.C. Wang Prize in Molecular Parasitology. Total amount: US\$2,000 (~ €1,848). (2019). Awarded for excellent performance at the Biology of Parasitism Course at the Marine Biological Laboratory, Woods Hole, MA, USA.
9. Funder: Organising Committee of the annual Sir William Dunn School of Pathology, University of Oxford. Total amount: £75 (~ €87). (2019). Best Poster in the Postdoc Symposium, Oxford, UK.

#### ORGANIZATION OF SEMINARS AND WORKSHOPS

**Symposium** “Leishmaniasis: What is it and why should Nepal care?”, 2023, Research Institute for Bioscience and Biotechnology, Kathmandu, Nepal, 20 attending participants, (1 day)

**Hands-on workshop** “Setting up a viability assay for eukaryotic organisms”, Research Institute for Bioscience and Biotechnology, Kathmandu, Nepal, 2023, 20 attending participants, (5 days)

*Responsibility:* Communication with sponsors, speakers, participant registration, chairing, curation of content, preparation of program, materials, advertisement, training and local technical assistance.

**Webinar** “Gene editing and regulation of protozoan parasites and their vectors”, 2022, 500 international attending participants, (2 days)

*Responsibility:* Communication with sponsors, speakers, participant registration, chairing, preparation of certificates, flyers, advertisement and local technical assistance.

**Hands-on workshop** “Gene editing in *Leishmania* parasites”, Instituto de Higiene e Medicina Tropical, Universidade NOVA de Lisboa, Portugal, 16 attending participants, (3 days)

**Responsibility:** Communication with sponsors, speakers, participant registration, chairing, teaching, preparation of certificates, flyers, advertisement, logistics and local technical assistance.

**Seminar** “Leishmaniasis: From Clinics to the laboratory”, 2013, Institute of Hygiene and Tropical Medicine – New University of Lisbon, Portugal, 72 attending participants, (2 hours)

**Responsibility:** Co-coordination of participant applications, preparation of certificates, flyers, logistics and technical assistance.

**Workshop** “Collection and fixation of tissues for morphological studies”, 2013, Institute of Hygiene and Tropical Medicine – New University of Lisbon, Portugal, 26 attending participants, (2 hours)

**Responsibility:** Preparation of logistics, reagents and materials for the execution of the workshop.

**Seminar** “Leishmaniasis in Portugal in the XXI century”, 2011, School of Veterinary Sciences of the University of Trás-os-Montes e Alto Douro, Alto Douro, Portugal, 242 attending participants, (3 hours)

**Responsibility:** Co-coordination of participant applications, preparation of certificates, flyers, logistics and technical assistance.

## SUPERVISING ACTIVITIES

*I am co-supervising the following undergraduate and graduate student projects and trainings with Prof. Dr. Pascal Mäser at Swiss TPH/Faculty of Science, University of Basel, Switzerland*

Michela Pirelli. **2025. MSc** in Infection Biology. “Establishing a sand fly and honeybee drug screening assay and barcoding *Leishmania* parasites”. (Ongoing until February 2026)

Betina Chalethu. **2025.** Voluntary internship. “Transformation of *E. coli*, plasmid amplification and pDNA quality control, for genetic manipulation of *Leishmania* sp.” (2 weeks)

*I co-supervised the following undergraduate and graduate student projects and trainings with Prof. Dr. Eva Gluenz in the United Kingdom and Switzerland*

### Student projects

Dana Geissbühler. **2024. BSc** Biology. “The role of the *Leishmania mexicana* P-type H<sup>+</sup> ATPase in pH regulation”

**Host institution:** Institute of Cell Biology, University of Bern, Switzerland

Romina Kalbermatten. **2024. BSc** Biology. “Localisation of *Leishmania mexicana* ABC transporter I3”

**Host institution:** Institute of Cell Biology, University of Bern, Switzerland

Sandro Möri. **2024. MSc** Biology. “The role of membrane transporter proteins in drug susceptibility in *Leishmania mexicana*”

**Host institution:** Institute of Cell Biology, University of Bern, Switzerland

Cagla Alagoz. **2023. PhD** Cell Biology. “Survival in the Macrophage: The Function of V-ATPase in the Infectivity of *L. mexicana* in the mammalian host”

**Host institution:** Institute of Cell Biology, University of Bern, Switzerland

Sandro Möri. **2023. BSc** Biology. “Susceptibility to Miltefosine and Amphotericin B in *Leishmania mexicana* mutants”

**Host institution:** Institute of Cell Biology, University of Bern, Switzerland

Robyn Chisholm. **2022. MSc** Infection Biology. “How do Protozoan Parasites utilise calcium?”

**Host institution:** Wellcome Centre for Integrative Parasitology, University of Glasgow, United Kingdom

George Dobson. **2022. MSc** Life Sciences. “Characterisation of *Leishmania* membrane transporters required for survival in macrophages”

**Host institution:** Wellcome Centre for Integrative Parasitology, University of Glasgow, United Kingdom

Caroline Ricce-Espada. **2019. PhD** Science. “Adjusting the CRISPR/Cas9 system to *Leishmania braziliensis* and study of the role of the Miltefosine transporter and Ros3 in *Leishmania* sp.”.

**Host institution:** Sir William Dunn School of Pathology, University of Oxford, United Kingdom

### Training internships

Ana Nunes. **2022. MSc** Biomedical Sciences. “Defining the localisation of calcium transporter proteins in *Leishmania mexicana* parasites”

**Host institution:** Wellcome Centre for Integrative Parasitology, University of Glasgow, United Kingdom

Olympia Tziouvara. **2022. PhD** Cellular Biology. “Lipid-Like Phosphatases of *Leishmania donovani*”

**Host institution:** Wellcome Centre for Integrative Parasitology, University of Glasgow, United Kingdom

Tomas Becvar. **2021. PhD** Parasitology. “Addback generation of double knock out *L. mexicana* cell line and cloning training.”

**Host institution:** Wellcome Centre for Integrative Parasitology, University of Glasgow, United Kingdom

*I co-supervised the following undergraduate and graduate student projects and trainings with Prof. Dr. Françoise Routier at the Institute of Cellular Chemistry, Department of Clinical Biochemistry, Medical School of Hannover, Germany.*

### Student projects

Markus Hinze. **2018. MD/PhD** Promotion. “Knock-down of DPY19 in *Toxoplasma gondii* tachizoites”.

Steffen Winkler. **2016. MSc** Biomembranes. “Optimizing *Caenorhabditis elegans* DPY-19 Immunostaining in *Drosophila* Schneider 2 cells.”

### Training internships

Caroline Meinshausen. **2017. Highschool** finalist. Internship for Parasitology.

Sebastian Böhlen. **2017. MSc** Biochemistry. Internship for Glycobiology.

Johannes Greve. **2016. MSc** Biochemistry. Internship for Glycobiology.

Leonie Ineundorf. **2015. MSc** Biochemistry. Internship for Glycobiology.

*I co-supervised the following undergraduate student projects and visiting researcher trainings with Dr. Sofia Cortes at the Institute of Tropical Hygiene and Medicine, NOVA University of Lisbon, Portugal.*

### Student projects

Liliana Lopes. **2014.** “Leishmanicidal effect of synthetic endoperoxides”, **BSc** Biomedical Sciences, University of Algarve, PT.

Romário Morais. **2014.** “Different *Leishmania* species DNA search in biological samples” **Bsc** Biology, Instituto Superior de Agronomia, PT.

Marisa Coelho. **2014.** “Optimization of an *in vitro* model of infection and recovery of *Leishmania* parasites for use in susceptibility studies”, **BSc** Applied Chemistry, Biotechnology, FCT-UNL, PT.

Tiago Nunes. **2013.** “Susceptibility of different strains of *Leishmania* to 1<sup>st</sup> line drug – Amphotericin B.” **BSc** Applied Chemistry, Biotechnology, FCT-UNL, PT.

Joana Marques. **2013.** “Impact of continuous axenic cultivation in *Leishmania infantum*.” **BSc** Applied Chemistry, Biotechnology, FCT-UNL, PT.

Geraldina Manjate. **2013.** “Evaluation of cell permissiveness to infection by strains of *Leishmania* spp..” **BSc** Biochemistry, FCT-UNL, PT.

### Training internships

Isabel Fragoso. **2014. MSc** Medicine, University of Algarve, PT.

Talita Nayara. **2013. BSc** Veterinary Medicine, Universidade Federal Rural de Pernambuco, BZ.

Liliana Lopes. **2013. BSc** Biomedical Sciences, Universidade do Algarve, PT.

Imen Bassoumi Jamoussi. **2013. MSc** Genetics, Instituto Pasteur de Tunis, TN.

**ACADEMIC TEACHING****Since 2022**

UC Molecular Parasitology (PT: Parasitologia Molecular), MSc Biomedical Sciences, “*Edição génica em tripanossomatídeos*”, IHMT-UNL (PT)

**Since 2021**

UCO Investigation in Leishmaniasis and applied methodologies (PT: Investigação em Leishmanioses e metodologias aplicadas), MSc Medical Parasitology, “*Edição génica em tripanossomatídeos*”, IHMT-UNL (PT)

UC Parasitology topics (PT: Tópicos em Parasitologia), MSc Pharmaceutical Sciences, “*Edição génica em tripanossomatídeos*”, Universidade Federal do Rio Grande do Norte (BZ)

**Since 2020**

UCO Cell Culture, MSc Biomedical Sciences, “*Utilização de linhas celulares para o cultivo de protozoários intracelulares obrigatórios: Toxoplasma e Leishmania*”, IHMT-UNL (PT)

**2013**

UCO *Leishmania* Molecular typing, PhD Biomedical Sciences (help in preparation and demonstration of practical classes), IHMT-UNL (PT)

UCO Leishmaniasis Module, MSc Medical Parasitology (help in preparation and demonstration of practical classes), IHMT-UNL (PT)

**PUBLIC ENGAGEMENT** (Last 5 years)**2024**

Speaker representing the Gluenz lab (IZB, UniBe) at the Biology day, “Now it is there, now it is not. How gene editing helps us discover the magical tricks of *Leishmania* parasites”, Lecture Hall of the Botanical Garden, Bern (CH)

**2023**

Science demonstrator representing the WCIP at the [Glasgow Science Festival](#), “*Can you find a cure for Leishmaniasis*”, Glasgow (UK)

Science demonstrator representing the WCIP at the Glasgow Science Centre, “*Science Lates: Meet the Expert*”, Glasgow (UK)

Consultant in the generation of a comic book on Leishmaniasis in [English](#) and [Portuguese](#) (a collaborative project between the WCIP PE office (Hannah Bialic) at the UofG and comic book artist Edward Ross, [www.edwardross.co.uk](http://www.edwardross.co.uk), from the [WCIP parasite comic series](#)), Glasgow (UK)

**2022**

Science demonstrator representing the WCIP at the [New Scientist Live](#), “*Neglected tropical diseases and where to find them*”, London (UK)

Creative director in the construction of a 3D-printed model of a female sand fly (a collaborative project between the WCIP PE office (Hannah Bialic) and the CAE and Graphics engineering department (Ewan Russel) at the UofG), Glasgow (UK)

Science demonstrator at the Einstein Garden “Parasites on tour!”, Greenman Festival, Wales (UK)

Research pitch talk at the [LUSO 2022](#), London (UK)

**2021**

Consultant in the generation of a Leishmaniasis and *Leishmania* transporter proteins related [animation](#)

**2020**

Creative director in the generation of a *Toxoplasma* and Glycobiology related [animation](#)

**ARTICLE REVISION BY INVITATION**

Emerging Infectious Diseases

Frontiers Cellular and Infection Microbiology

Frontiers Molecular Biosciences, Parasitology

Gene

International Journal for Parasitology: Parasites and Wildlife

Molecular Microbiology  
Parasite & Vectors  
Preventive Veterinary Medicine

#### **MEMBERSHIP OF PROFESSIONAL SOCIETIES**

Swiss Society for Tropical Medicine and Parasitology (since June 2024)

Microbiology Society (since April 2021)

British Society for Cell Biology (since February 2020)

Biochemical Society (since September 2018)

PARSUK (since September 2018)

Marie Curie Alumni Association (since July 2014)

British Society of Parasitology (since September 2012)